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Optimizing Bacterial Adhesion to a Microfluidic Platform for Monitoring Bacterial Biofilm Growth

Aaron Cheng

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Mentors: Mariana Meyer, Peter Dykstra

Professor Reza Ghodssi



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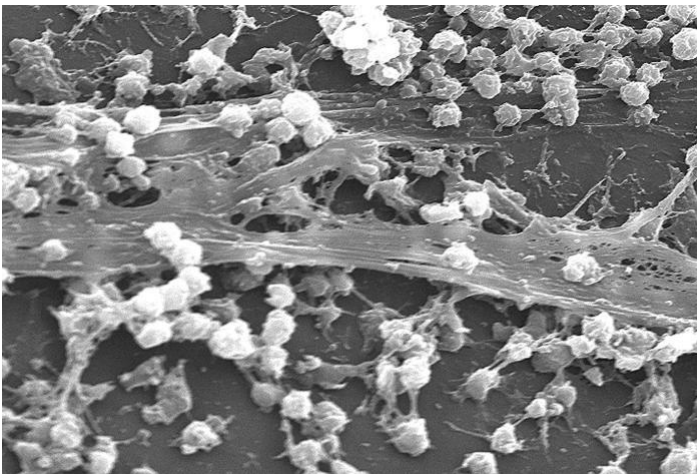
Bacterial Quorum Sensing

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- Quorum Sensing
 - Cell-cell communication mechanism for unicellular organisms
 - Uptake/release of chemical signaling molecules
 - 3 principal classes of signaling molecules:
 - *E. coli.* : autoinducer-2 (AI-2) - “universal” between Gram-positive and Gram-negative
- Changes in gene expression if bacterial population threshold reached
 - Ex. production of biofilm matrix, toxins

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- Biofilms:
 - Composed primarily of polysaccharides, protein, DNA, and bacterial cells
 - Prevalent in infections (respiratory tract, urinary tract) and foreign implants (heart valves, joint prostheses)
 - Dense matrix - increased antibiotic resistance



Biofilm on inserted catheter, www.cdc.gov

QuickTime™ and a
decompressor
are needed to see this picture.

3D image of bacteria adhesion (confocal microscopy)



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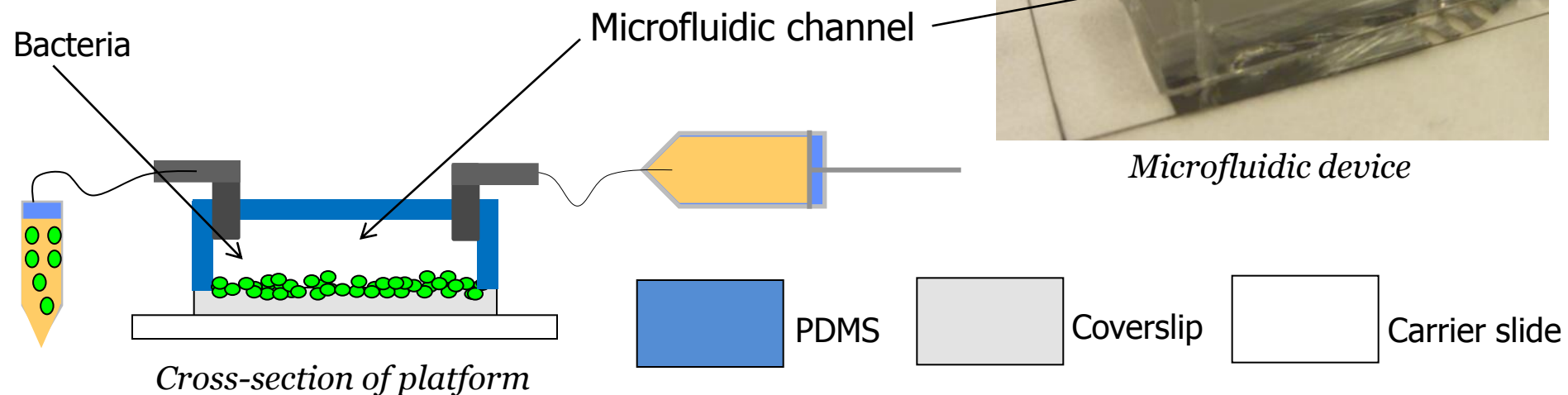
Studying Biofilms

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- Real-time optical monitoring of biofilm growth in microfluidics
- Advantages:
 - Highly controllable environment
 - Inexpensive, fast setup
 - Parallel operation
- Poly-L-lysine and fibrinogen shown to increase bacterial adhesion in generic bacteria - may increase repeatability of biofilm formation

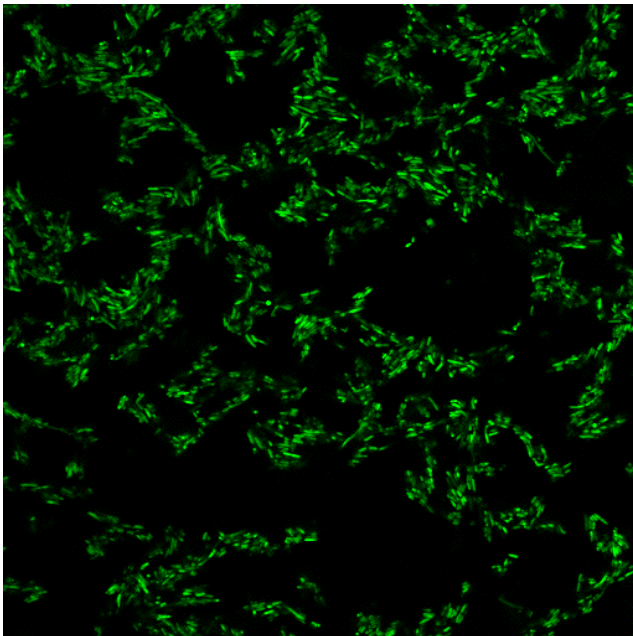
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- A biocompatible transparent polymer (PDMS) used to create microfluidic channel
- Device sterilized, inoculated with bacteria, rinsed with LB growth media

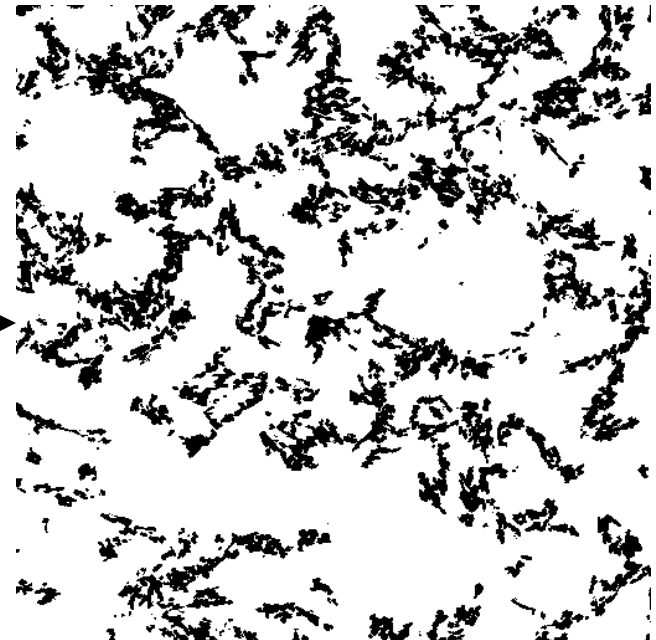


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- Image bacteria in the channel using fluorescence microscopy
- Images analyzed using ImageJ to calculate percent area covered by bacteria



Fluorescence image

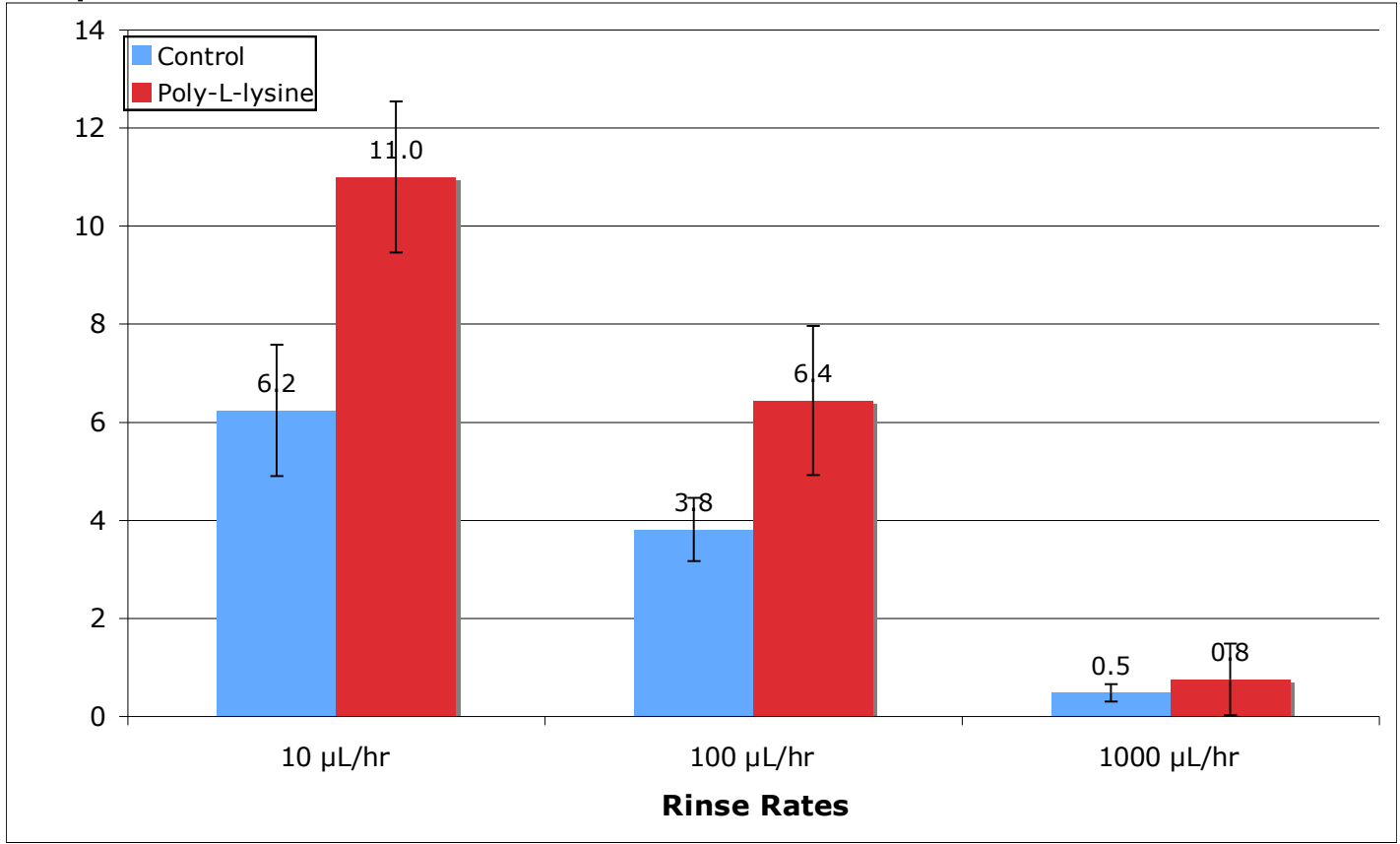


ImageJ analysis



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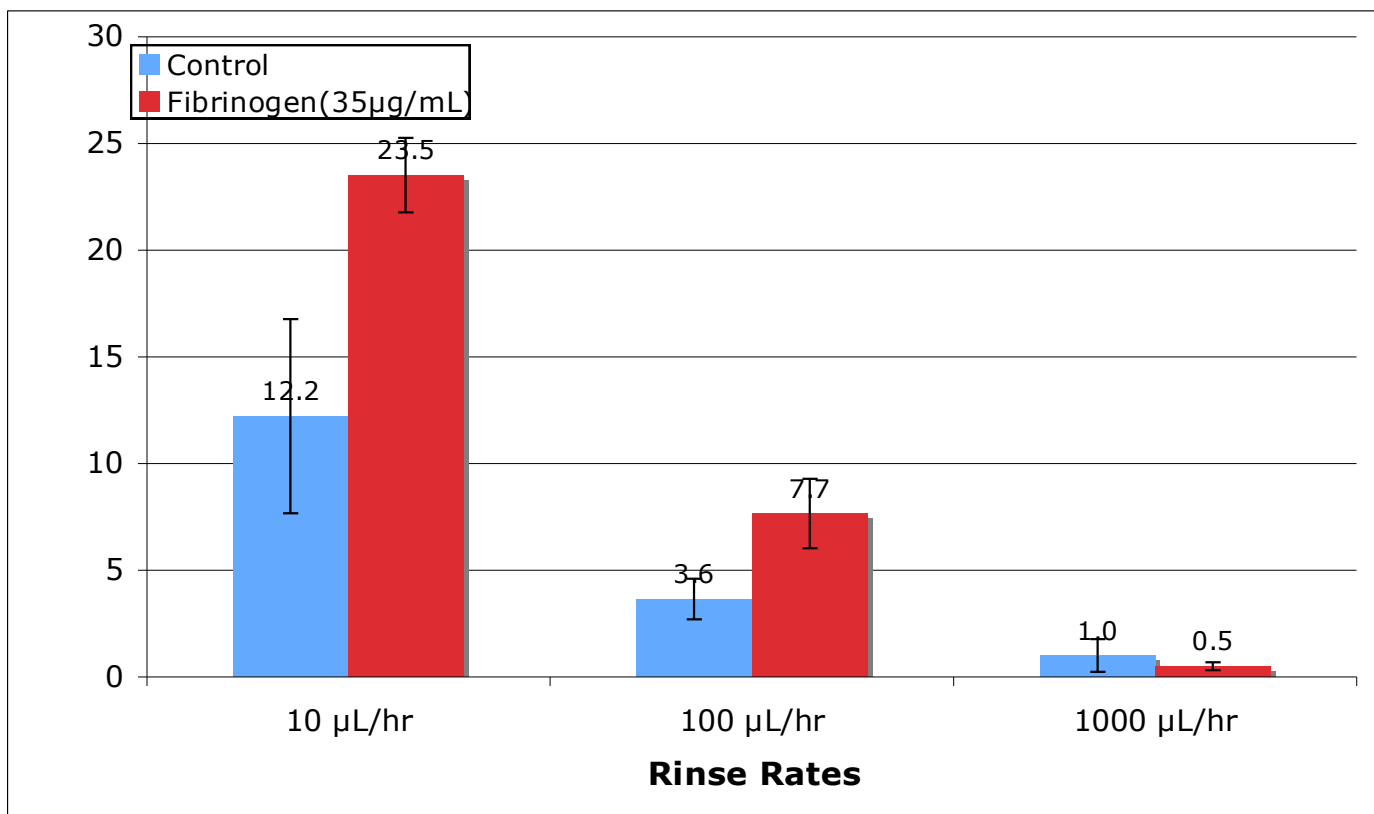
- PLL coated channels vs. uncoated channels subjected to different rinse rates
- 0.01% PLL flow into channel, dried over night
- Bacteria of OD₆₀₀ 1.2, incubated 2hrs at 37 °C, rinsed with LB growth media 30min at 10μL/hr





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- FG coated channels vs. uncoated channels subjected to different rinse rates
- 35 μ g/mL FG in PBS flowed into channel, incubated 1hr at 37 °C, rinsed with LB growth media 30min at 50 μ L/hr
- Bacteria of OD₆₀₀ 1.2, incubated 2hrs at 37 °C, rinsed with LB growth media 30min at 10 μ L/hr

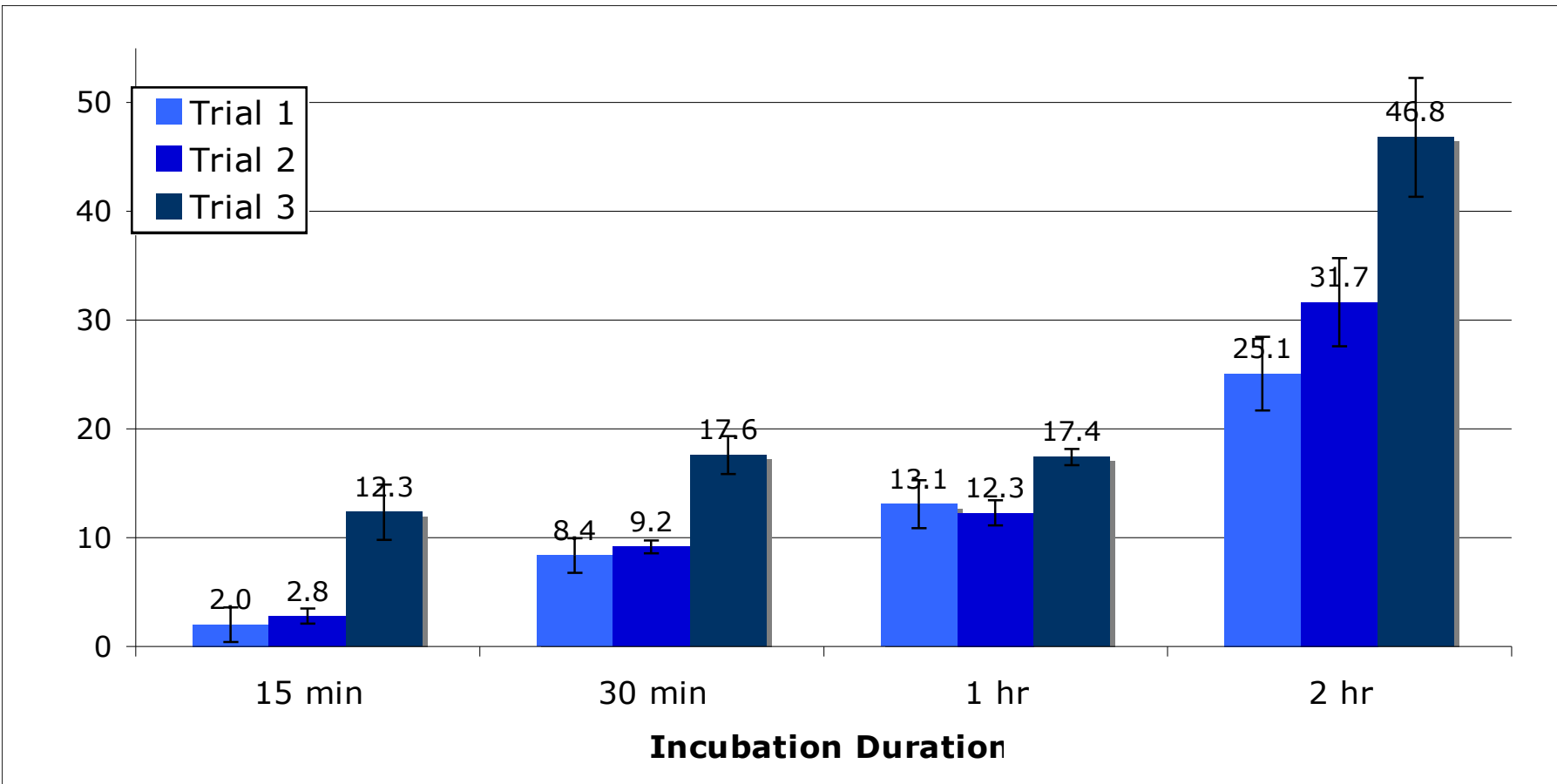




Results with Varying Incubation Times

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- Varying bacterial incubation times with uncoated channels
- Bacteria of OD₆₀₀ 1.3, incubated at 37 °C, rinsed with LB growth media 30min at 10 μL/hr





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- Poly-L-lysine and fibrinogen shown to improve bacterial adhesion to the microfluidic device
- Bacterial adhesion decreases as flow rate of the rinsing procedure increases
- The longer the incubation time, the more bacteria attach to the surface of the device
- Incorporation of the results will likely improve bacterial biofilm repeatability in the microfluidic device



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Acknowledgments

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